

ANTIMICROBIAL ACTIVITY OF THREE MEDICINAL PLANTS

Nizar Sirag^{1,*}, Sitelbanat Yassin², Mirghani A Yousif²

1- Department of Chemistry and Pharmacognosy, Faculty of Pharmacy, University of Gezira, Sudan.

2- Department of Pharmaceutics, Faculty of Pharmacy, University of Gezira, Sudan.

* Corresponding author.

الخلاصة

تستخدم النباتات الطبية تقليديا لعلاج بعض الأمراض المسببة بواسطة الجراثيم . الغرض من هذه الدراسة اختبار ثلاثة أنواع من النباتات مستخدمه في الطب الشعبي كمضادات حيوية وقورنت مع المواد المرجعية. أظهرت المستخلصات الكحولية لهذه النباتات نتائج متفاوتة. وجد أن للعفص نشاطا عاليا ضد الايشريكيه القولونية بينما ابدى مستخلص عرق السوس نشاطا ملحوظا علي الزائفة الزنجارية, كذلك نتج اثر تثبيطي علي الايشريكيه القولونية عند استخدام حب الرشاد. يمتلك العفص وحب الرشاد أثرا واضحا علي فطر المبيضة البيضاء بينما افتقر عرق السوس هذا الأثر.

يمكن أن نستنتج أن الثلاث نباتات المختبرة تعتبر مصادر متوقعة لمضادات حيوية جديدة ولذلك يرجى عمل بحوث أخرى لفصل المواد المسؤولة عن النشاط المضاد للميكروبات.

ABSTRACT

Background: Medicinal plants have been traditionally used for different kinds of ailments including infectious diseases.

Objectives: The present work was undertaken to investigate the antimicrobial activity of three plants used in folk medicine.

Methods: Plant extracts of (*Quercus infectoria*, *Glycyrrhiza glabra* and *Lepidium virginicum*) were *in vitro* tested against four bacterial strains and one type of fungi for their antimicrobial activities by using well diffusion techniques.

Results: The ethanolic extracts of these plants showed varying results. *Quercus infectoria* was found to possess the highly marked antibacterial activity against both *E. coli* and *Staphylococcus aureus* while *Glycyrrhiza glabra* exhibited a significant antibacterial effect against *Pseudomonas aurginosa*. *Lepidium*

EDITORIAL

virginicum produced antibacterial activity against *E.coli*. Both *Quercus infectoria* and *Lepidium virginicum* caused a considerable antifungal effect, while *Glycyrrhiza glabra* devoid of such antifungal activity.

Conclusion: The three tested plant extracts could be considered as potential sources of new antimicrobial agents. Further researches should be made to isolate compounds responsible for antimicrobial activities.

Key words: Antimicrobial activity, Medicinal plants, Folk medicine.

INTRODUCTION

Some bacteria and fungi are extremely pathogenic causing serious human infections. The discovery of antibiotics to combat these pathogens marked a resolution in the 20th century (1). Unfortunately, because of the inappropriate use of antibiotics in human, certain strains of bacteria and fungi developed the ability to produce substances which block the action of antimicrobial agents or change their target or ability to penetrate cells (2, 3). To substitute synthetic agents, many of today modern and effective drugs have their origin in traditional Folk Medicine (4). Plants have been used to treat human, animals, and plant diseases from time immemorial. Also herbal medicines have been known to man for centuries (5, 6). Therapeutic efficacy of many indigenous plants for many disorders has been described by practitioners of traditional medicine (7–9).

The discovery of medicinal plants in different parts of the world is important to medicine sector, helping in establishment of new directions towards propagation of alternative medicinal plants that offer safe, effective, non toxic, better economic and social benefits (10).

MATERIALS AND METHODS

Plants materials:

Plants used for this study were purchased from the local market. All plant samples were taxonomically identified by the Department of Chemistry and Pharmacognosy, Faculty of Pharmacy, University of Gezira (Table 1).

Microorganism used:

The following strains of bacteria were used: *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Staphylococcus aureus* and *Escherichia coli*. The yeast strain used in this study was *Candida albicans*. All microorganisms used in the present study were previously identified and kindly supplied by Medical Laboratory, University of Gezira.

Table 1: Plants screened for antimicrobial activities

Botanical name/ family	Common/vernacular name	Plant part tested
1. <i>Quercus infectoria</i> Family: Fagaceae	Gall nut (Afas)	Fruit
2. <i>Glycyrrhiza glabra</i> Family: leguminosae	Liquorice root (Irgsoos)	Root
3. <i>Lepidium virginicum</i> Family: Brassicaceae	Hab-Elrashad	Seed

Extraction of plant materials

This was carried out as described by Doughari *et al.*, 2007. Twenty grams of the coarsely powdered plant samples were extracted separately by maceration, using ethanol, in a conical flask for 72 hours, filtered and dried at room temperature and kept in a refrigerator until use. Working solutions were then made for each plant extract by using distilled water to have the corresponding concentrations of 1 gram (plant material)/ml.

In vitro testing of extracts for antimicrobial activity:

Testing for antibacterial activity:

The cup-plate agar diffusion method was adopted according to (12) to assess the antibacterial activity of the prepared ethanolic extracts (1gm/ml). Microorganisms were sub-cultured from slope agar into nutrient agar, incubated at 37 °C for 18 hours in the incubator. Wells were made at the center of each of the four different sectors of the plate, three of them were filled with 100 µl of plant extract and the other was filled with 100µl of the control (Ceftriaxone 0.1gm). Each extract was tested against organisms in four replicates. Plates were then left for 2 hours at room temperature and then incubated at 37 °C for 18 hours. The inhibition zones were observed and the results were interpreted by measuring the diameter of clear zones of inhibition (Table 2).

Testing for antifungal activity:

The same method as for bacteria was adopted. Instead of nutrient agar, yeast and mould extract agar was used. The inoculated medium was incubated at 25 °C for 48 hours for *Candida albicans*. Nystatin the commonly used antifungal agent was used as control (100,000 I.U). Results were displayed in Table2.

RESULTS AND DISCUSSION

The *in vitro* antibacterial and antifungal activities of various morphological parts of the three ethanolic plant extracts against selected microorganisms are shown in Table 2.

It was found that the three plant extracts produced a considerable antifungal

EDITORIAL

activity against the tested species except *Glycyrrhiza glabra* which produced no effect against *Candida albicans*.

High antibacterial activity was produced by *Quercus infectoria* against *E. coli*, followed by *Glycyrrhiza glabra* against *pseudomonas aueruginosa* and the least antibacterial effect was elicited by *Glycyrrhiza glabra* against *E. coli*.

These results coincide with those reported in literature (13 – 16).

Furthermore, this study revealed that *Quercus infectoria* and *Lepidium virginicum* extracts were found to possess remarkable inhibitory effects on both bacterial and fungal growth that make such plant species to be considered as a promising antimicrobial agent (17 – 19).

Quercus infectoria gave negative antibacterial effect against *Pseudomonas aueruginosa* as well as *Glycyrrhiza glabra* that lacks anticandidal effect in contrast to data reported in literature(1) and as claimed by practice. These varying results can be attributed to the differences in methods of extraction and biological variations in species used.

Table 2: Antimicrobial activity of the tested plants

Plant Extracts (Plant material) I gm/ml	Mean diameter of clear zone of inhibition in (mm)				
	<i>Pseudomonas aurginosa</i>	<i>Klebsiella pneumoniae</i>	<i>Staphylococcus aureus</i>	<i>E.coli</i>	<i>Candida albicans</i>
<i>Quercus infectoria</i>	-	15	35	37	22.7
<i>Glycyrrhiza glabra</i>	25.7	14.7	15.7	4.3	-ve
<i>Lepidium virginicum</i>	16	15	28	6.3	19.7
Control Cefterioxone 0.1g/ml	21	16.7	45	20	-
Nystatin 100000IU/ml	-	-	-	-	25

CONCLUSION

The three tested plant extracts could be considered as potential sources of new antimicrobial agents. Further researches should be made to isolate compounds responsible for antimicrobial activities.

ACKNOWLEDGEMENT

The authors would like to acknowledge the positive contribution of the following students: Mohamed BadrEldin, Sawsan A.Elateef and Zenab Omer.

REFERENCES

1. Evans, C.W (1992). Trease and Evans. Pharmacognosy, (13 edition) Bailliere Tindall, London 758-762.
2. Ali, B.H; Bashir, A.K, and Tanira, M.O.H (1995). Anti-inflammatory, antipyretic and analgesic effects of *Lawsonia inermis* in rats. Pharmacology 51:356-363.
3. Boji, H; Steven, J; Kenneth, A and Bachmann, A (2005). A nationwide survey on bacterial respiratory infections in ambulatory patients. *Journal of clinical epidemiology*. 58:414-420.
4. Natarajan, V; Venugopal, PV, and Menon, T (2003). Effect of *Azadirachta indica* on the growth pattern

EDITORIAL

- of dermatophytes. *Indian journal of medicine and microbiology*. 21:98-101.
5. Goun, E; Cunningham, D; Chu, C and Mile, D (2003). Antibacterial and antifungal activity of Indonesian ethno medical plants. *Fitoterapia*.74:592-596.
 6. Misra, SK and Sahu, HC (1977). Screening of some indigenous plants for antifungal activity against dermatophytes. *Indian journal of pharmacology* 9:269-272.
 7. Almaqbool, AZ; Bashir, AK and Salih, AKM (1988). Antimicrobial activity of certain Sudanese plants used in Folkloric Medicine. *Fitoterapia* 56:59-62.
 7. Iqbal, Z; Shabeen, H and Sheraz, B (2002). Antifungal properties of some indigenous plants from Peshawar Valley. *Asian journal of plant science* .1:708-709.
 8. Khattak, SG; Gilani, SN and Ikram, M (1985). Antipyretic studies on some indigenous Pakistani medicinal plants. *Journal of ethnopharmacology* .14:45-52.
 9. Elie, K; Vatche, K; Rbih, S and Salma, N. (2004). Screening of selected indigenous plants of Lebanon for antimicrobial activity. *Journal of ethnopharmacology*. 93:1-7.
 10. Doughari, J.H; Pukuma, M.S and De, N (2007). Antibacterial effects of *Balanites aegyptica* and *Moringa oleifera* on Salmonella typhi. *African journal of Biotechnology*. 6(19)2212-2215.
 11. Jinga, P; Nehal, K and Sumitra, C (2006). Evaluation of antibacterial activity and phytochemical analysis of *Bauhinia variiegata* bark. *Journal of biomedical research* .9(1)53-56.
 12. Samo Andresek; Breda Simvnovska and Irena Vovk (2004). Antimicrobial and Antioxidative enrichment of Oak (*Quercus robur*), *International Journal of Food Microbiology*: 92; 181- 187.
 13. Ayfer, S and Turgey, A (2003). Antimicrobial activities of various medicinal and commercial plant extracts. Department of Biology, Faculty of Arts and Science, Imam University, Kahramanmaras, Turkey.
 14. Gulluce, M; Sengul, M and Karaman, I (2008). Antimicrobial effect of *Quercus*. *Journal of Phytotherapy Research*: 18: (3): 208-211.
 15. Vivek , K; Gupta; Atiya Fatima ; Uzma , F (2008). Antimicrobial activity of *Glycyrrhiza glabra* root. *Journal of Ethnopharmacology*: 116: (2): 377-380.
 16. Gislene, G.F; Nascimento, J.L and Silva, L (2000). Antibacterial activity of plant extracts and phytochemicals on antibiotic resistant bacteria. *Brazilian journal of microbiology* 31:247-256.
 17. Bassari, D.F and Fan, S.H (2004). The potential of aqueous and acetone extracts of galls of *Quercus infectoria* as antibacterial agents. *Indian journal of pharmacology* 37(1):26-29.
 18. Supayang, P; Sasitorn, C and Sakol, S (2008). *Quercus infectoria*. *Pharmaceutical biology*. 46(6):367-372